

TITLE: Plating, Passaging and Freezing of Fibroblasts and Keratinocytes

SOP Number: D-DEZ-PRO-002

Revision Number: 0

Effective Date: 07 Apr 2015

2. Dump contents of biopsy sample vial onto petri dish. Rinse sample in biopsy rinsing solution. Transfer biopsy to a 6-well TC-treated plate and mince into fine pieces using a sterile scalpel.
3. Allow pieces to air dry ~5 mins. Carefully add 3 mL of biopsy media to each well.
4. Label plate with correct biopsy number, date, and scientist's initials.
5. Place plate in incubator. Incubator settings: 5.0% CO₂, 37 degrees Celsius and 20% O₂.
6. Feed cells Monday, Wednesday, and Friday with 3 mL of biopsy media per well.

F. PASSAGING/FREEZING FIBROBLASTS

7. When cells are confluent you can either passage or freeze depending on downstream application, i.e. reprogramming or long-term storage.
8. To passage or freeze add 1 mL TrypLE Express to each well. Incubate for 5 minutes at 37 degrees Celsius. Agitate the cells and pipette into a 15 mL conical tube with 3 mL of Biopsy media.
9. Centrifuge for 5 minutes at room temperature at 200g.
10. Aspirate supernatant leaving only the cell pellet.
11. To passage, re-suspend pellet in Biopsy media at a density of 100,000 cells/mL. Plate in 6-well TC-treated plate at a density of 250,000 cells per well. Label as indicated above with addition of passage number, i.e. P1. Feed as described above until cells are confluent and ready to be passaged and re-plated or cryo-protected and frozen.
12. To freeze, re-suspend pellet in 0.5 mL of Biopsy media/500,000 cells and add 0.5 mL to each cryo-vial. Add 0.5 mL Freezing media to each cryo-vial. Label cryo-vials with biopsy number, passage number, date and Scientist's initials. Place vials in Mr. Freeze and place in -80 degrees Celsius freezer overnight. 24 hrs later remove vials from Mr. Freeze and place immediately into liquid nitrogen for long-term storage.

I. History

Effective Date	Revision	Change
07 Apr 2015	0	Initial issue of SOP